



*California Environmental Protection Agency*

**AIR RESOURCES BOARD**

## Method 424

# Determination of Inorganic Cadmium Emissions From Stationary Sources

Adopted: January 22, 1987

## Method 424 – Determination of Inorganic Cadmium Emissions From Stationary Sources

### 1. APPLICABILITY AND PRINCIPLE

#### 1.1 Applicability

This method applies to the measurement of inorganic cadmium emissions from stationary sources.

#### 1.2 Principle

Particulate and gaseous emissions are extracted isokinetically from the stack and passed through an impinger-filter train where the cadmium is collected on a glass-fiber filter and in a solution of nitric acid. The combined filter extract and impinger solution are analyzed for cadmium by atomic absorption spectrometry (direct aspiration method).

### 2. RANGE AND SENSITIVITY

#### 2.1 Range

Detection limits, sensitivity and optimum concentration ranges will vary with the make and model of the atomic absorption spectrophotometer. However, when a wavelength of 228.8 nm is used, the optimum concentration range is typically 0.05 to 2 mg/L, and the detection limit is 0.005 mg/L.

#### 2.2 Analytical sensitivity

The typical sensitivity for this method is 0.25 mg/L.

### 3. INTERFERENCES

3.1 Nonspecific absorption and light scattering can be significant at the analytical wavelength. Therefore background correction is necessary.

3.2 Samples and standards should be monitored for viscosity differences that may alter the aspiration rate.

3.3 Solution matrix effects can be kept to a minimum with the Method of Standard Additions.

If the analytical method herein recommended does not give the desired sensitivity in the presence of interfering substances in the sample, the tester may select an equivalent procedure, subject to approval by the Executive Officer. The

tester must then produce data to demonstrate that the method is equivalent, and substantiate this data through an adequate quality assurance program approved by the Executive Officer.

#### 4. APPARATUS

The following sampling apparatus is recommended. The tester may use alternative sampling apparatus only if, after review by the Executive Officer, it is deemed equivalent for the purposes of this test method.

##### 4.1 Sampling Train

A schematic diagram of the sampling train is shown in Figure 1. This is similar to the ARB Method 5 with some minor changes which are described below.

4.1.1 Probe Nozzle, Probe Liner, Pitot Tube, Differential Pressure Gauge, Filter Holder, Filter Heating System, Metering System, Barometer and Gas Density Determination Equipment. Same as Method 5, Sections 2.1.1 to 2.1.6, and 2.1.8 to 2.1.10, respectively.

4.1.2 Impingers. Four impingers are connected in series with glass ball joint fittings. The first, third, and fourth impingers are of the Greenburg-Smith design modified by replacing the tip with a 1-cm (0.5 in.) I.D. glass tube extending to 1 cm from the bottom of the flask. The second impinger is of the Greenburg-Smith design with the standard tip.

The first and second impingers shall contain known quantities of 0.1 N HNO<sub>3</sub> (Section 6.1.3). The third shall be empty, and the fourth shall contain a known weight of silica gel or equivalent desiccant.

A thermometer which measures temperatures to within 1°C (2°F), should be placed at the outlet of the fourth impinger.

##### 4.2 Sample Recovery.

The following items are needed:

4.2.1 Probe Liner and Probe Nozzle Brushes, Petri Dishes, Plastic Storage Containers, Rubber Policeman and Funnel. Same as Method 5, Sections 2.2.1, 2.2.4, 2.2.6, and 2.2.7, respectively.

4.2.2 Wash Bottles. Glass (2)

4.2.3 Sample Storage Containers. Chemically resistant, borosilicate glass bottles, for 0.1 nitric acid ( $\text{HNO}_3$ ) impinger and probe solutions and washes, 1000 mL. Use screw-cap liners that are either rubber-backed Teflon or leak-free and resistant to chemical attack by 0.1 N  $\text{HNO}_3$  (Narrow mouth glass bottles have been found to be less prone to leakage).

4.2.4 Graduated Cylinder and/or Balance. To measure condensed water to within 2 mL or 1 g. Use a graduated cylinder that has a minimum capacity of 500 mL, and subdivisions no greater than 5 mL. (Most laboratory balances are capable of weighing to the nearest 0.5 g or less).

4.2.5 Funnel. Glass, to aid in sample recovery.

#### 4.3 Analysis

The following equipment is needed:

4.3.1 Atomic Absorption Spectrophotometer with an air-acetylene burner head and cadmium hollow cathode lamp.

4.3.2 Strip chart recorder (optional).

4.3.3 Hot Plate.

4.3.4 Erlenmeyer Flasks. 125 mL. 24/40  $\bar{\text{F}}$ .

4.3.5 Whatman No. 42 filter paper (or equivalent).

4.3.6 Volumetric Flasks. 100-mL, 200-mL, 250-mL, and 1000-mL.

### 5. REAGENTS

Unless otherwise specified, use American Chemical Society reagent grade (or equivalent) chemicals throughout. Mention of trade names or specific products does not constitute endorsement by the California Air Resources Board.

#### 5.1 Sampling.

The following reagents are needed:

5.1.1 Filters, Silica Gel, Crushed Ice and Stopcock Grease. Same as Method 5, Sections 3.1.1, 3.1.2, 3.1.4 and 3.1.5, respectively.

- 5.1.2 Water. Deionized, distilled, to conform to ASTM Specification D1193-77. Type 3 (Reference 10.1). If high concentrations of organic matter are not expected to be present, the analyst may omit the potassium permanganate test for oxidizable organic matter.
- 5.1.3 Nitric Acid, 0.1 N. Dilute 6.5 mL of concentrated HNO<sub>3</sub> to 1 liter with deionized distilled water (It may be desirable to run blanks before field use so that all analyses can be corrected).
- 5.2 Pretest Preparation. 6 N HNO<sub>3</sub> is needed. Dilute 390 mL of concentrated HNO<sub>3</sub> to 1 liter with deionized distilled water.
- 5.3 Sample Recovery. 0.1 N HNO<sub>3</sub> (same as 5.1.4 above).
- 5.4 Analysis. The following reagents are needed (use ACS reagent grade chemicals or equivalent, unless otherwise specified):
- 5.4.1 Water. Same as 5.1.3 above.
- 5.4.2 Nitric Acid. Concentrated. Acid should be analyzed to determine whether the sample analysis should be corrected.
- 5.4.3 Nitric Acid, 50 percent (v/v). Dilute 500 mL of concentrated HNO<sub>3</sub> to 1 liter with deionized distilled water.
- 5.4.4 Cadmium standard stock solution (1000 mg Cd/L). This may be a commercially available standard that has been certified, and is traceable to NBS standards, or it may be prepared as follows:
- Dissolve 1.000 g of spectroscopically pure Cadmium metal in a minimum volume of (1+1) HCl. Dilute to 1 liter with 1% (v/v) HCl (1 ml = 1 mg Cd). THIS METAL IS TOXIC, AND SHOULD BE HANDLED WITH EXTRA CARE.
- 5.4.5 Intermediate Stock Standards. Pipet 2 mL of the stock Cadmium standard solution (5.4.4) into a 200-mL volumetric flask, and dilute to volume with deionized distilled water. This corresponds to 10 mg Cd/L.
- 5.4.6 Working Cadmium Standards. Pipet 0.0, 1.0, 2.0, 3.0, 4.0 and 5.0 mL of the intermediate stock standard (5.4.5) into 200-mL volumetric flasks. Add 2 mL of concentrated HNO<sub>3</sub> to each flask and dilute to volume with deionized distilled water. The concentrations of these working standards are 0.0, 0.05, 0.75, 1.00, 1.50, and 2.00 mg Cd/L, respectively. Prepare additional standards

at other concentrations as needed. The final concentration of acid in the working standards should be the same as that of the sample to be analyzed.

- 5.4.7 Air. Suitable quality for atomic absorption analysis. The air should be cleaned, dried, and filtered to remove oil, water and other foreign substances. The air may be supplied from a compressed air line, a laboratory compressor, or a cylinder of compressed air, or as recommended by the instrument manufacturer.
- 5.4.8 Acetylene. Suitable quality for atomic absorption analysis. Commercial grade acetylene or as recommended by the instrument manufacturer. Acetone, which is usually present in acetylene cylinders, can be prevented from entering and affecting flame conditions by replacing the cylinder before the pressure has fallen to 50 psig.
- 5.4.9 Hydrogen peroxide, 3 percent (v/v). Dilute 10 mL of 30 percent  $\text{H}_2\text{O}_2$  to 100 mL with deionized distilled water.

## 6. PROCEDURE

### 6.1 Sampling.

Because of the complexity of this method, testers should be trained and experienced with the test procedures in order to ensure reliable results.

- 6.1.1 Pretest Preparation. Follow the same general procedure given in Method 5 Section 4.1.1, except the filter need not be weighed.
- 6.1.2 Preliminary Determinations. Follow the same general procedure given in Method 5, Section 4.1.2.
- 6.1.3 Preparation of Collection Train. Follow the same general procedure given in Method 5, Section 4.1.3, except place 100 mL of 0.1 N  $\text{HNO}_3$  in each of the first two impingers, leave the third impinger empty, and transfer approximately 200 to 300 g of preweighed silica gel from its container to the fourth impinger. Assemble the train as shown in Figure 1.
- 6.1.4 Leak-Check Procedures. Follow the general leak-check procedures given in Method 5, Sections 4.1.4.1 (Pretest Leak Check), 4.1.4.2 (Leak-Checks During the Sample Run), and 4.1.4.3 (Post-Test Leak-Check).

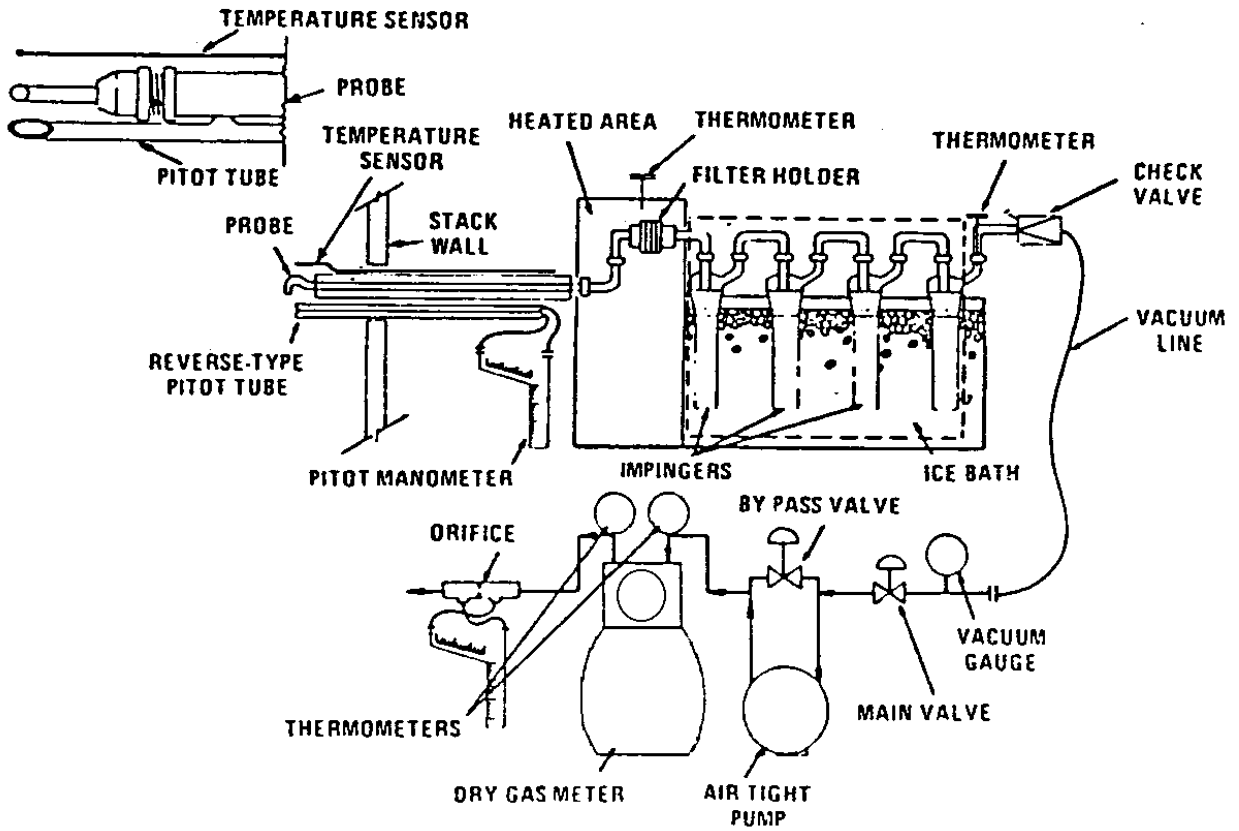


Figure 1 Inorganic Cadmium Sampling Train

6.1.5 Sampling Train Operation. Follow the same general procedure given in Method 5, Section 4.1.5. For each run, record the data required on a data sheet such as the one shown in CARB Method 5, Figure 5-2.

6.1.6 Calculation of Percent Isokinetic. Same as Method 5, Section 4.1.6.

## 6.2 Sample Recovery.

Begin proper clean-up procedure as soon as the probe is removed from the stack at the end of the sampling period.

Allow the probe to cool. When it can be safely handled, wipe off all external particulate matter near the tip of the probe nozzle and place a cap over it. Do not cap off the probe tip tightly while the sampling train is cooling down as this would create a vacuum in the filter holder, thus drawing liquid from the impingers into the filter.

Before moving the sampling train to the cleanup site, remove the probe from the sampling train, wipe off the silicone grease, and cap the open outlet of the probe. Be careful not to lose any condensate that might be present. Wipe off the silicone grease from the glassware inlet where the probe was fastened and cap the inlet. Remove the umbilical cord from the last impinger and cap the impinger. The tester may use ground-glass stoppers, plastic caps, or serum caps to close these openings.

Transfer the probe and filter-impinger assembly to a cleanup area, which is clean and protected from the wind so that the chances of contaminating or losing the sample are minimized.

Inspect the train prior to and during disassembly and note any abnormal conditions. Treat the samples as follows:

6.2.1 Container No. 1 (Filter): Carefully remove the filter from the filter holder and place it in its identified petri dish container. If it is necessary to fold the filter, make sure that the sample-exposed side is inside the fold. Carefully transfer to the petri dish any visible sample matter and/or filter fibers that adhere to the filter holder gasket using a dry Nylon bristle brush and/or sharp-edged blade. Seal the container.

6.2.2 Container No. 2 (Probe). Taking care that dust on the outside of the probe or other exterior surfaces does not get into the sample, clean all surfaces that have been exposed to the sample (including



the probe nozzle, probe fitting, probe liner, and front half of the filter holder) with 0.1 N HNO<sub>3</sub>. Place the wash in a glass sample storage container. Measure and record (to the nearest 2-mL) the total amount of 0.1 N HNO<sub>3</sub> used for each rinse. Perform the rinses with 0.1 N HNO<sub>3</sub> as follows:

Carefully remove the probe nozzle and rinse the inside surface with 0.1 N HNO<sub>3</sub> from a wash bottle. Brush with a nylon-bristle brush, and rinse until the 0.1 N HNO<sub>3</sub> rinse shows no visible particles, after which, make a final rinse of the inside surface. Brush and rinse the inside parts of the Swagelok fitting with 0.1 N HNO<sub>3</sub> in a like manner until no visible particles remain.

Rinse the probe liner with 0.1 N HNO<sub>3</sub>. While squirting the nitric acid rinse into the upper end of the probe, tilt and rotate the probe so that all inside surfaces are wetted with the nitric acid. Let the 0.1 N HNO<sub>3</sub> drain from the lower end into the sample container. The tester may use a glass funnel to aid in transferring liquid washes to the container. Follow the rinse with a probe brush. Hold the probe in an inclined position, and squirt 0.1 N HNO<sub>3</sub> into the upper end as the probe brush is being pushed with a twisting action through the probe. Hold the sample container underneath the lower end of the probe, and catch any liquid and particulate matter brushed from the probe. Run the brush through the probe three times or more until no visible sample matter is carried out with the 0.1 N HNO<sub>3</sub> and none remains on the probe liner on visual inspection. With stainless steel or other metal probes, run the brush through in the above prescribed manner at least six times, since metal probes have small crevices in which particulate matter can be entrapped. Rinse the brush with 0.1 N HNO<sub>3</sub> and quantitatively collect these washings in the sample container. After the brushing, make a final rinse of the probe as described above. It is recommended that two people clean the probe to minimize loss of sample: Between sampling runs, keep brushes clean and protection from contamination.

After ensuring that all joints have been wiped clean of silicone grease, brush and rinse with 0.1 N HNO<sub>3</sub> the inside of the front half of the filter holder. Brush and rinse each surface three times or more, if needed, to remove visible particulate matter. Make a final rinse of the brush and filter holder. After all 0.1 N HNO<sub>3</sub> washings and particulate matter have been collected in the sample container, tighten the lid on the sample container so that the liquid will not leak out when it is shipped to the laboratory. Mark the height of the fluid level to determine whether leakage occurs during transport. Label the container clearly to identify its contents.

Rinse the glassware a final time with water to remove residual  $\text{HNO}_3$  before reassembling. Do not save the final rinse water.

6.2.3 Container No. 3 (Silica Gel). Check the color of the indicating silica gel to determine if it has been completely spent and note its condition. Transfer the silica gel from the fourth impinger to the original container and seal. The tester may use a funnel to pour the silica gel and rubber policeman to remove the silica gel from the impinger. It is not necessary to remove the small amount of particles that may adhere to the impinger walls and are difficult to remove. Since the gain in weight is to be used for moisture calculations, do not use any water or other liquids to transfer the silica gel. If a balance is available in the field, the tester may follow procedure for Container No. 3 under Section 6.4 (Analysis).

6.2.4 Container No. 4 (Impingers). If the volume of liquid is large, the tester may place the impinger solutions in several containers. Clean each of the first three impingers and connecting glassware in the following manner:

1. Wipe the impinger ball joints free of silicone grease and cap the joints.
2. Rotate and agitate each impinger, so that the impinger contents might serve as a rinse solution.
3. Transfer the contents of the impingers to a 500-mL graduated cylinder. Remove the outlet ball joint cap and drain the contents through this opening. Do not separate the impinger parts (inner and outer tubes) while transferring their contents to the cylinder. Measure the liquid volume to within  $\pm 2$  mL. Alternatively, determine the weight of the liquid to within  $\pm 0.5$  g. Record in the log the volume or weight of the liquid present, along with a notation of any color or film observed in the impinger catch. The liquid volume or weight is needed, along with the silica gel data, to calculate the stack gas moisture content (see Method 5, Figure 5-3).
4. Transfer the contents to Container No. 4.

NOTE: In steps 5 and 6 below, measure and record the total amount of 0.1 N  $\text{HNO}_3$  used for rinsing.

5. Pour approximately 30 mL of 0.1 N  $\text{HNO}_3$  into each of the first three impingers and agitate the impingers. Drain the 0.1 N  $\text{HNO}_3$  through the outlet arm of each impinger into

Container No. 4. Repeat this operation a second time; inspect the impingers for any abnormal conditions.

6. Wipe the ball joints of the glassware connecting the impingers free of silicone grease and rinse each piece of glassware twice with 0.1 N HNO<sub>3</sub>; transfer this rinse into Container No. 4. (Do not rinse or brush the glass-fritted filter support.) Mark the height of the fluid level to determine whether leakage occurs during transport. Label the container to clearly identify its contents.

6.2.5 Blanks. Save 200 mL of the 0.1 N HNO<sub>3</sub> used for sampling and cleanup as a blank. Take the solution directly from the bottle being used and place into a glass sample container.

### 6.3 Sample Preparation.

6.3.1 Container No. 1 (Filter). Cut the filter into strips and transfer the strips and all loose particulate matter into a 125-mL Erlenmeyer flask. Rinse the petri dish with 10 mL of 50 percent HNO<sub>3</sub> to insure a quantitative transfer and add to the flask. (Note: If the total volume required in Section 5.3.3 is expected to exceed 80 mL, use a 250-mL Erlenmeyer flask in place of the 125 mL flask.

6.3.2 Containers No. 2 and No. 4 (Probe and Impingers). Check the liquid level in Containers No. 2 and/or No. 4 and to determine whether any sample was lost during shipment. Record observations on the analysis sheet. If a noticeable amount of leakage has occurred, either void the sample or take steps, subject to the approval of the Executive Officer to adjust the final results. Combine the contents of Containers No. 2 and No. 4 and take to dryness on a hot plate.

6.3.3 Sample Extraction for Cadmium. Using the approximate stack gas particulate concentration and the total volume of stack gas sampled, estimate the total weight of particulate sample collected. Then transfer the residue from containers No. 2 and No. 4 to the 125-mL Erlenmeyer flask that contains the filter using a rubber policeman and 10 mL of 50 percent HNO<sub>3</sub> for every 100 mg of sample collected in the train or a minimum of 30 mL of 50 percent HNO<sub>3</sub> whichever is larger.

Place the Erlenmeyer flask on a hot plate and heat with periodic stirring for 30 min at a temperature just below boiling. If the sample volume falls below 15 mL, add more 50 percent HNO<sub>3</sub>. Add 10 mL of 3 percent H<sub>2</sub>O<sub>2</sub> and continue heating for 10 min. Add 50 mL of

hot (80°C) deionized distilled water and heat for 20 min. Remove the flask from the hot plate and allow to cool. Filter the sample through Whatman No. 42 filter paper (or equivalent) and transfer the filtrate to a 250-mL volumetric flask. Dilute to volume with deionized distilled water. The diluted sample has an approximate acid concentration of 3% (v/v) HNO<sub>3</sub> and is now ready for analysis.

- 6.3.4 Filter Blank. Determine a filter blank using two filters from each lot of filters used in the sampling train. Cut each filter into strips and place each filter in a separate 125-mL Erlenmeyer flask. Add 15 mL of 50 percent HNO<sub>3</sub> and treat as described in Section 6.3.3 using 10 mL of 3 percent H<sub>2</sub>O<sub>2</sub> and 50 mL of hot, deionized distilled water. Filter and dilute to a total volume of 100 mL using deionized distilled water.
- 6.3.5 0.1 N HNO<sub>3</sub> Blank. Take the entire 200 mL of 0.1 N HNO<sub>3</sub> to dryness on a steam bath, add 15 mL of 50 percent HNO<sub>3</sub>, and treat as described in Section 6.3.3 using 10 mL of 3 percent H<sub>2</sub>O<sub>2</sub> and 50 mL of hot, deionized distilled water. Dilute to a total volume of 100 mL using deionized distilled water.

#### 6.4 Analysis.

- 6.4.1 Cadmium Determination. Calibrate the spectrophotometer as described in Section 7.2. Aspirate each sample, the filter blank, and 0.1 N HNO<sub>3</sub> blank and determine the concentration either directly or from the calibration curve and determine the absorbance for each source sample, the filter blank, and 0.1 N HNO<sub>3</sub> blank. Analyze each sample three times in this manner. Use a minimum of one blank per sample batch to determine if contamination or memory effects are present. If the absorbance values for the samples are above the linear range of the standards, dilute the solutions with 0.1 N HNO<sub>3</sub>, reanalyze and use the appropriate dilution factor in calculations. If the Cd concentration of a sample is at the low end of the calibration curve and high accuracy is required, the sample can be taken to dryness on a hot plate and the residue dissolved in the appropriate volume of water to bring it into the optimum range of the calibration curve.
- 6.4.2 Mandatory Check for Matrix Effects. The analysis for Cd by atomic absorption is sensitive to the chemical composition and to the physical properties (viscosity, pH) of the sample (matrix effects). Since the Cd procedure described here will be applied to many different sources, many sample matrices will be encountered. Therefore, check (mandatory) at least one sample from each source using the Method of Additions to ascertain that the chemical

composition and physical properties of the sample have not caused erroneous analytical results.

Three acceptable "Method of Additions" procedures are described in the General Procedure Section of the Perkin Elmer Corporation Manual (Reference 10.1). If the results of the Method of Additions procedure on the source sample do not agree within 5 percent of the value obtained by the conventional atomic absorption analysis, then the tester must reanalyze all samples from the source using the Method of Additions procedure.

6.4.3 Container No. 3 (Silica Gel). The tester may conduct this step in the field. Weigh the spent silica gel (or silica gel plus impinger) to the nearest 0.5 g; record this weight.

## 7. CALIBRATION.

### 7.1 Sampling Train Calibration.

Calibrate the sampling train components according to the indicated sections of Method 5: Probe Nozzle (Section 5.1); Pitot Tube (Section 5.2); Metering System (Section 5.3); Probe Heater (Section 5.4); Temperature Gauges (Section 5.5); Leak-Check of the Metering System (Section 5.6); and Barometer (Section 5.7).

### 7.2 Spectrophotometer.

Use the 228.8-nm wavelength line and background correction. An oxidizing air-acetylene flame is required. Follow the manufacturer's operating instructions for all other instrument parameters.

Measure the absorbance of the standard solutions. Repeat until good agreement ( $\pm 3$  percent) is obtained between two consecutive readings. For those instruments which read directly in concentrations, set the curve corrector to read out the proper concentration.

Construct a calibration curve by plotting the absorbance of the standards (y-axis) versus concentration in  $\mu\text{g Cd/mL}$  (x-axis), or by using the methods of standard additions if necessary. Draw or compute a straight line through the linear portion of the curve. Do not force the calibration curve through zero, but if the curve does not pass through the origin or at least lie closer to the origin than  $\pm 0.003$  absorbance units, check for incorrectly prepared standards and for curvature in the calibration curve.

To determine stability of the calibration curve, run a blank and a standard after every five samples and recalibrate, as necessary. Calibration curves must be composed of a minimum of a blank and three standards

Spiked samples or standard reference materials should be run periodically to ensure that correct procedures are being followed, and that all equipment are operating properly.

## 8. CALCULATIONS.

### 8.1 Dry Gas Volume.

Using the data from this test, calculate  $V_{m(\text{std})}$  the total volume of dry gas metered corrected to standard conditions (20° and 760 mm Hg), by using Equation 5-1 of Method 5. If necessary, adjust  $V_{m(\text{std})}$  for leakages as outlined in Section 6.3 of Method 5. Check the field data sheet for the average dry gas meter temperature and average orifice pressure drop.

### 8.2 Volume of Water Vapor and Moisture content.

Using data obtained in this test and Equations 5-2 and 5-3 of Method 5, calculate the volume of water vapor  $V_{w(\text{std})}$  and the moisture content  $B_{ws}$  of the stack gas.

### 8.3 Total Cadmium in Source Sample.

For each source sample correct the average absorbance for the contribution of the filter blank and the 0.1 N HNO<sub>3</sub> blank. Use the calibration curve and this corrected absorbance to determine the Cd concentration in the sample aspirated into the spectrophotometer. Calculate the total Cd content  $C^{\circ}_{Cd}$  (in ug) in the original source sample. Correct for all the dilutions that were made to bring the Cd concentration of the sample into the linear range of the spectrophotometer.

### 8.4 Cadmium Concentration.

Calculate the stack gas Cadmium concentration  $C_{Cd}$  in mg/dscm as follows:

$$C_{Cd} = \frac{K C^{\circ}_{Cd}}{V_{m(\text{std})}} \quad \text{Eq. 1}$$

Where:

K = 0.001 mg/ug for metric units.  
= 2.205 x 10<sup>-9</sup> lb/ug for English units.

8.5 Isokinetic Variation and Acceptable Results.

Same as Method 5, Sections 6.11 and 6.12, respectively. To calculate  $V_s$ , the average stack gas velocity, use equation 2-9 of Method 2 and the data from this field test.

9. ALTERNATIVE TEST METHODS FOR INORGANIC CADMIUM.

9.1 Method 213.2, issued in 1978, is an EPA method that has been approved for NPDES and SDWA. The analytical technique is flameless atomic absorption spectrophotometry. The optimum concentration range is typically 0.5 – 10 ug/L and the detection limit is 0.1 ug/L. Hence, this method is recommended for measurement of levels of cadmium below 20 ug/L. Intralaboratory and interlaboratory studies have demonstrated that the method will yield results with acceptable precision, accuracy, and specificity (Reference 10.7).

9.2 Method 6010 (Reference 10.5), and NIOSH Method 7300 (Reference 10.8). These methods utilize Inductively Coupled Plasma Atomic Emission Spectroscopy (ICP). This is an analytical technique that requires much skill to interpret results and obtain valid data.

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- 10.8 NIOSH Manual of Analytical Methods, 3<sup>rd</sup> ed., Method 7048, Cadmium and compounds, as Cd (Atomic Absorption, Flame) U.S. Department of Health and Human Services DHHS (NIOSH) Publ. No 84-100. Feb, 1984.
- 10.9 Same as Method 5, Citations 2 to 5 and 7 of Section 7.